## Three Unusual New Sesquiterpenes from Alpinia oxyphylla

by Jun-Ju Xu<sup>a</sup>)<sup>b</sup>), Ning-Hua Tan\*<sup>a</sup>), Yi-Shan Chen<sup>a</sup>)<sup>b</sup>), Xu-Lin Pan<sup>a</sup>), Guang-Zhi Zeng<sup>a</sup>), Hong-Jin Han<sup>a</sup>), Chang-Jiu Ji<sup>a</sup>)<sup>b</sup>), and Mei-Ju Zhu<sup>a</sup>)

a) State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, P. R. China

(phone: +86-871-5223800; fax: +86-871-5223800; e-mail: nhtan@mail.kib.ac.cn)

b) Graduate University of the Chinese Academy of Sciences, Beijing 100049, P. R. China

Two new norcadinene sesquiterpenes oxyphyllones C and D (1 and 2, resp.), and one new 1,10-secognaiane sesquiterpene, (+)-mandassidion (3), together with two known compounds, oxyphyllenodiol B (4) and (1R,2R)-p-menth-3-ene-1,2-diol (5), were isolated from the fruits of Alpinia oxyphylla. Their structures were determined on the basis of spectroscopic analysis, including 2D-NMR spectroscopic techniques. Compounds 1, 2, and 3 exhibited no cytotoxicity against three cancer cell lines.

Introduction. – The plant of *Alpinia oxyphylla* Miq. (Zingibereae) is widely distributed in South China and used as a folk medicine to treat intestinal disorders, urosis, diuresis, ulceration, and dementia [1–3]. Previous phytochemical investigations of this plant have led to the isolation of bioactive compounds including many sesquiterpenes [1][2]. In our present research, three unusual new sesquiterpenes including two 14-norcadinene sesquiterpenes oxyphyllones C (1) and D (2), and a 1,10-secognaiane sesquiterpene, (+)-mandassidion (3), together with two known compounds, oxyphyllenodiol B (4) [1] and (1*R*,2*R*)-*p*-menth-3-ene-1,2-diol (5) [4][5], were isolated from the fruits of *A. oxyphylla*. Compounds 1, 2, and 3 were tested for their activities towards human-tumor A549, HT-29, and SGC-7901 cell lines, none of them showed cytotoxicities in these assays at 10 μg/ml according to the method described in [6]. In this article, we mainly report the isolation and the structure elucidation of the new sesquiterpenes.

**Results and Discussion.** – Oxyphyllone C (1) was obtained as a colorless oil. The molecular formula  $C_{14}H_{20}O_2$  with five degrees of unsaturation was deduced on the basis of HR-ESI-MS (m/z 243.1357 ([M+Na] $^+$ ; calc. 243.1360)). The  $^1H$ -NMR data (Table) showed three Me signals at  $\delta(H)$  0.99 (d, J=6.8, Me(12)), 0.91 (d, J=6.8, Me(13)), and 1.48 (s, Me(15)), three CH signals at  $\delta(H)$  3.18 (s, H-C(4)), 2.30–2.36 (m, H-C(6))

and 1.99–2.15 (m, H–C(11)), and four CH<sub>2</sub> signals. The <sup>13</sup>C-NMR and DEPT spectra revealed the presence of an  $\alpha$ , $\beta$ -conjugated ketone system, resonating at  $\delta$ (C) 197.5 (s, C(9)), 155.2 (s, C(5)), and 133.9 (s, C(10)), respectively, and an epoxy linkage between C(3) ( $\delta$ (C) 63.3 (s)) and C(4) ( $\delta$ (C) 57.9 (d)). The above information suggested that compound 1 was an unusual degraded cadinene sesquiterpene, similar to the known compound oxyphyllenodiol B [1].

Table. <sup>1</sup> H- and <sup>13</sup> C-NMR Data of	f <b>1</b> and <b>2</b> at 400/100 MHz.	Respectively. In CDCl <sub>3</sub> ; $\delta$ in ppm,	J in Hz.

	1		2	
	¹H	<sup>13</sup> C	¹H	<sup>13</sup> C
1	2.65 (br. $dd$ , $J = 5.9$ , 16.9, $H_a$ ), 1.82–1.96 $(m, H_{\beta})$	18.1 (t)	7.88 (d, J = 8.0)	127.4 (d)
2	$1.50-1.55 \ (m, H_a), 1.99-2.15 \ (m, H_{\beta})$	26.8(t)	7.19 (d, J = 8.0)	128.8(d)
3		63.3(s)		144.1 (s)
4	3.18 (s)	57.9 (d)	7.43 (br. <i>s</i> )	126.2(d)
5		155.2(s)		148.7 (s)
6	2.30-2.36 (m)	44.8(d)		73.7(s)
7	$1.82-1.96 \ (m, H_a), 1.99-2.15 \ (m, H_{\beta})$	23.3 (t)	2.04-2.16 (m), 2.36-2.42 (m)	33.0(t)
8	$2.45-2.52 (m, H_a), 2.30-2.36 (m, H_b)$	34.5 (t)	2.63-2.79 (m)	34.6 (t)
9		197.5(s)		197.4 (s)
10		133.9(s)		128.8 (s)
11	1.99-2.15 (m)	29.9(d)	2.04-2.16 (m)	34.9(d)
12	0.99 (d, J = 6.8)	21.3(q)	1.01 (d, J = 6.8)	17.5(q)
13	0.91 (d, J = 6.8)	19.5(q)	0.93 (d, J = 6.8)	16.3 (q)
15	1.48 (s)	21.5 (q)	2.43 (s)	44.0 (q)

The planar structure of **1** was determined by  ${}^{1}H$ ,  ${}^{1}H$ -COSY and HMBC correlations (*Fig. 1*). In the  ${}^{1}H$ ,  ${}^{1}H$ -COSY spectrum, two spin systems corresponding to  $CH_2(1)$ /  $CH_2(2)$  and to Me(12), and Me(13)/H-C(11)/H- $C(6)/CH_2(7)/CH_2(8)$  were observed, which were extended by HMBC correlations of  $CH_2(1)$  with C(9) and C(10), of Me(15) with C(2), C(3), and C(4), of H-C(4) with C(5), C(6) and C(10), of H-C(6) with C(5) and C(10), and  $CH_2(8)$  with C(9).

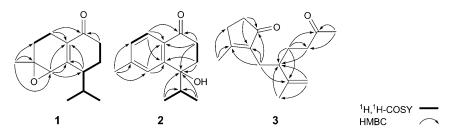


Fig. 1. <sup>1</sup>H, <sup>1</sup>H-COSY and key HMBC for compounds **1-3** 

From the key NOESY correlations (Fig. 2) between H–C(4) and Me(12) ( $\delta$ (H) 0.99) and H $_{\beta}$ –C(6) ( $\delta$ (H) 2.30–2.36), and by comparing with the data of oxyphyllenodiol B, the Me–C(3) and iPr–C(6) groups should be in  $\beta$ - and  $\alpha$ -orientation,

respectively [1], which was further supported by the computer-generated (Gaussian 03, B3LYP/6-31G\*\*) interatomic distance of H-C(4) to H-C(6), 2.41~Å < 3~Å [7][8]. However, when the Me-C(3) and iPr-C(6) groups were fixed in  $\alpha$ -orientation, the calculated interatomic distance of H-C(4) and H-C(6) was 3.53~Å > 3~Å. Moreover, from the biosynthetic point of view, the co-occurrence of compounds 1 and oxyphyllenodiol B in the same plant suggested that oxyphyllenodiol B could be the precursor of 1, which was in agreement with the above deduction. Thus, the relative configuration of 1 was established.

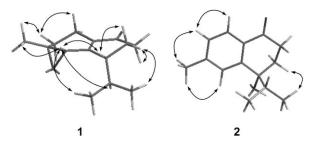


Fig. 2. Key ROESY correlations for 1 and 2

Oxyphyllone D (2) was obtained as a colorless, optically inactive oil. Its HR-ESI-MS (m/z 241.1206 ([M+Na]<sup>+</sup>)) showed the molecular formula  $C_{14}H_{18}O_2$ . <sup>13</sup>C-NMR and DEPT spectra (Table) indicated 14 C-atom signals due to seven sp<sup>2</sup> and seven sp<sup>3</sup> C-atoms (5 C, 4 CH, 2 CH<sub>2</sub>, 3 Me). Moreover, the seven sp<sup>2</sup> C-atom signals were ascribed to a CO and a 1,3,4-trisubstituted aromatic ring, which was further confirmed by the <sup>1</sup>H-NMR spectrum signals at  $\delta$ (H) 7.88 (d, J=8.0, 1 H), 7.19 (d, J=8.0, 1 H), and 7.43 (br. s, 1 H). According to the <sup>1</sup>H, <sup>1</sup>H-COSY, key HMBC and NOESY correlations, a norcadinene-type skeleton of 2 was established ( $Figs.\ 1$  and 2), with the OH group located at C(6), based on the HMBC correlations: the cross-peaks from Me(12) and Me(13) to C(6), from CH<sub>2</sub>(7) and CH<sub>2</sub>(8) to C(6) and with a C(9)=O group, derived from the HMBC correlations: the cross-peaks from H–C(1) and CH<sub>2</sub>(8) to C(9). Since this compound which is similar to 4-hydroxy-4,7-dimethyl-1-tetralone [9][10], showed no optical activity, it must be a racemic mixture.

(+)-Mandassidion (3; ( $[\alpha]_D^{28} = +33.22$ ) was obtained as a colorless oil. The molecular formula of 3 was revealed as  $C_{15}H_{22}O_2$  by HR-ESI-MS (m/z 257.1517 ( $[M+Na]^+$ )), <sup>13</sup>C-NMR, and DEPT spectra. The NMR data as compared with literature suggested that compound 3 was a 1,10-secognaiane sesquiterpene with the same planar structure to mandassidion ( $[\alpha]_D = -9.1$ ) [11], which was supported by HSQC and key HMBC correlations (*Fig. 1*). Comparison of the optical value of mandassidion, compound 3 was determined as an enantiomer of mandassidion.

## **Experimental Part**

General. Column chromatography (CC): silica gel (SiO<sub>2</sub>; 100-200 or 200-300 mesh), and silica gel H (60  $\mu$ m), both obtained from *Qingdao Marine Chemical Co.*, P. R. China. TLC: silica gel GF254. Semiprep. reversed-phase (RP) HPLC: *Agilent 1100* liquid chromatograph, with a Zorbax  $SB-C_{18}$  column. MCI: CHP-20P. Sephadex: LH-20. Optical rotations:  $Jasco\ DIP-370$  digital polarimeter. UV

Spectra: Shimadzu 210A double-beam spectrophotometer;  $\lambda_{\max}$  (log  $\varepsilon$ ) in nm. IR Spectra: Bio-Rad FTS-135 spectrometer; KBr pellets; in cm<sup>-1</sup>. NMR Spectra: Bruker DRX-500 and Bruker AM-400 instruments, with Me<sub>4</sub>Si as internal standard;  $\delta$  in ppm. EI-MS: VG Auto Spec-3000 mass spectrometer; in m/z (rel. %). ESI and HR-ESI-MS: API Qstar Pulsar instrument.

Plant Material. The fruits of Alpinia oxyphylla were bought from Kunming medicinal market, Kunming, Yunnan Province, P. R. China and identified by Prof. Ning-Hua Tan.

Extraction and Isolation. The dried and powdered fruits of Alpinia oxyphylla (15 kg) were extracted with acetone/ $H_2O$  (70%) under reflux for 8 h (5 × 30 l). The resulting residue was partitioned between petroleum ether (PE) and  $H_2O$ , AcOEt and  $H_2O$ , and then BuOH and  $H_2O$ . The AcOEt extract (480 g) was separated by CC on SiO<sub>2</sub>, eluting with PE/acetone (9:1 to 1:1) to yield ten fractions (*Frs.* 1–10). *Fr.* 1 (16 g) was subjected to CC (SiO<sub>2</sub>, PE/CHCl<sub>3</sub>/acetone 3:1:0.1) to afford two subfractions (*Fr.* 1.1 and *Fr.* 1.2). *Fr.* 1.2 was further purified by repeated CC (*RP-18*) and HPLC (MeOH/ $H_2O$  6:4) to yield 1 (3 mg). *Fr.* 4 (49 g) was subjected to CC (SiO<sub>2</sub>, CHCl<sub>3</sub>/AcOEt, 100:1 to 9:1) to afford four subfractions (*Frs.* 4.1–4.4). *Fr.* 4.2 was further purified by repeated CC (*MCI*) and *Sephadex* (*LH-20*) and HPLC (MeCN/ $H_2O$  3:7) to yield 2 (3 mg) and 3 (24 mg). *Fr.* 8 (5 g) was subjected to CC (*RP-18*, MeOH/ $H_2O$  85:15 to 1:1) to afford three subfractions (*Frs.* 8.1–8.3). *Fr.* 8.3 was further purified by CC (SiO<sub>2</sub>, CHCl<sub>3</sub>/ $Me_2CO$  3:1 and PE/AcOEt 1:2) to yield 4 (10 mg) and 5 (10 mg).

Oxyphyllone C (=(1aR\*,7R\*,7bS\*)-1a,3,5,6,7,7b-Hexahydro-1a-methyl-7-(propan-2-yl)naph-tho[1,2-b]oxiren-4(2H)-one; 1). Colorless oil. [ $\alpha$ ] $_{\rm D}^{\rm 13}$  = -308.19 (c = 0.15, CHCl $_3$ ). UV (MeOH): 196 (3.53), 262 (3.94). IR (KBr): 3425, 2958, 2925, 1662.  $^{\rm 1}H$ - and  $^{\rm 13}$ C-NMR: Table. HR-ESI-MS: 243.1357 ([M+Na] $^+$ ,  $C_{14}H_{20}$ NaO $_2^+$ ; calc. 243.1360).

Oxyphyllone D (= 3,4-Dihydro-4-hydroxy-6-methyl-4-(propan-2-yl)-naphthalen-1(2H)-one; 2). Colorless oil. [ $\alpha$ ] $_{0}^{27}$  = 0 (c = 0.35, CHCl $_{3}$ ). UV (MeOH): 209 (4.19), 260 (4.00). IR (KBr): 3442, 2966, 1670, 1605.  $^{1}$ H- and  $^{13}$ C-NMR: Table. HR-ESI-MS: 241.1206 ([M + Na] $^{+}$ ,  $C_{14}H_{18}$ NaO $_{2}^{+}$ ; calc. 241.1204).

 $(+)\text{-}Mandassidion \ (=3\text{-}Methyl-2\text{-}[5\text{-}oxo\text{-}2\text{-}(prop\text{-}1\text{-}en\text{-}2\text{-}yl)hexyl]cyclopent\text{-}2\text{-}en\text{-}1\text{-}one; \ \textbf{3}). \ \text{Colorless oil.} \ [a]_{\text{D}}^{28} = +33.22 \ (c = 0.15, \text{CHCl}_3). \ \text{UV (MeOH): } 236 \ (3.72). \ ^1\text{H-NMR (CDCl}_3, 400 \ \text{MHz): } 4.67 \ \text{(br. } s, \ H_a - \text{C}(12)); \ 4.57 \ \text{(br. } s, \ H_b - \text{C}(12)); \ 2.45 - 2.48 \ (m, \ \text{CH}_2(2)); \ 2.30 - 2.40 \ (m, \ \text{CH}_2(3), \ \text{CH}_2(9)); \ 2.17 - 2.28 \ (m, \ \text{H} - \text{C}(7), \ \text{CH}_2(6)); \ 2.08 \ (s, \ \text{Me}(15)); \ 2.00 \ (s, \ \text{Me}(14)); \ 1.49 - 1.62 \ (m, \ \text{CH}_2(8)); \ 1.59 \ (s, \ \text{Me}(13)). \ ^{13}\text{C-NMR (CDCl}_3, \ 100 \ \text{MHz}): \ 209.5 \ (s, \ \text{C}(10)); \ 209.0 \ (s, \ \text{C}(1)); \ 171.0 \ (s, \ \text{C}(4)); \ 146.3 \ (s, \ \text{C}(11)); \ 138.8 \ (s, \ \text{C}(5)); \ 112.4 \ (t, \ \text{C}(12)); \ 45.1 \ (d, \ \text{C}(7)); \ 41.5 \ (t, \ \text{C}(9)); \ 34.2 \ (t, \ \text{C}(3)); \ 31.5 \ (t, \ \text{C}(2)); \ 30.0 \ (q, \ \text{C}(15)); \ 27.6 \ (t, \ \text{C}(6)); \ 26.2 \ (t, \ \text{C}(8)); \ 18.0 \ (q, \ \text{C}(13)); \ 17.5 \ (q, \ \text{C}(14)). \ \text{HR-ESI-MS: } 257.1517 \ ([M+Na]^+, \ \text{C}_{13}\text{H}_{22}\text{NaO}_2^+; \ \text{calc. } 257.1515). \$ 

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